

11/06/05

10/526741
JT12 Rec'd PCT/PTO 04 MAR 2005

DESCRIPTION

ANTIBODY AGAINST SECRETED N-TERMINAL PEPTIDE OF GPC3 PRESENT IN BLOOD OR C-TERMINAL PEPTIDE OF GPC3

Technical Field

The present invention relates to an antibody against an N-terminal peptide or C-terminal peptide of GPC3. More specifically, the invention relates to an antibody against a GPC3 N-terminal peptide of about 40 kDa as found in the soluble form of the GPC3 core protein. Additionally, the invention also relates to an antibody against a GPC3 C-terminal peptide of about 30 kDa as found in the soluble form of the GPC3 core protein.

Background Art

The presence of the glypican family is reported as a new family of heparan sulfate proteoglycan existing on cell surface. Up to now, it is reported that five types of glypican (glypican 1, glypican 2, glypican 3, glypican 4 and glypican 5) exist. The members of the family have a core protein of a uniform size (about 60 kDa) and have unique cysteine residues well conserved in common, and are bound to cell membrane via glycosylphosphatidylinositol (GPI) anchor.

Glypican 3 (GPC3) is known to be deeply involved in cell

division during development and the control of the pattern thereof. Additionally, it is known that the GPC3 gene is highly expressed in hepatoma cell and that the GPC3 gene is possibly used as a marker of hepatocellular carcinoma.

The present inventors previously found that an anti-GPC3 antibody had an ADCC activity and a CDC activity and was useful as the therapeutic treatment of hepatoma and filed a patent application (Japanese Patent Application 2001-189443).

However, GPC3 is a membrane-bound protein and it has not been reported that a GPC3 protein of secreted form existed. Thus, no examination has been made about the use of the GPC3 protein itself as a tumor marker in blood.

Disclosure of the Invention

The present inventors found a fact that glypican 3 (GPC3) is cleaved at an amino acid residue 358 thereof or at an amino acid residue 374 thereof or a region in the vicinity of the residues. On an assumption that the soluble form of GPC3 would be secreted in the blood of hepatoma patients, the inventors established a GPC3 sandwich ELISA system to show the existence of the secreted form of GPC3 in the culture supernatant of human hepatoma cell HepG2 highly expressing GPC3. Further, the inventors successfully assayed the secreted form of GPC3 not only in the plasma of a mouse transplanted with HepG2 but also in the serum of a human hepatoma patient. Because the expression

of the GPC3 gene is observed in hepatoma at an earlier stage compared with the time involving the occurrence of AFP as a hepatoma marker, the inventors considered that the detection of GPC3 would be useful for cancer diagnosis. Additionally because it appears to be hard to detect the secreted form of GPC3 with an anti-GPC3 antibody recognizing a C-terminal peptide fragment, the secreted form of GPC3 was assumed to be dominantly present as an N-terminal peptide fragment. Thus, the inventors considered that an anti-GPC3 antibody recognizing the N terminus was preferably used for detecting the secreted form of GPC3. Accordingly, the inventors made an attempt to develop an antibody recognizing the N-terminal peptide of GPC3, and thus have achieved the invention. Further, the inventors found that an antibody against the C terminus of GPC3 had a high cytotoxic activity and considered that the use of the anti-GPC3 antibody recognizing the C terminus would be preferable for disrupting cancer cell, i.e. for therapeutically treating cancer. Then, the inventors made an attempt of developing an antibody recognizing the C-terminal peptide of GPC3, and thus have achieved the invention.

Since it is observed that GPC3 is expressed in cancer cell lines other than hepatoma cell lines, such as lung cancer, colon cancer, breast cancer, prostate cancer, pancreatic cancer, and lymphoma, GPC3 may possibly be applied to the diagnosis of cancers other than hepatoma.

Specifically, the invention relates to an antibody against an N-terminal peptide of GPC3.

Additionally, the invention relates to the antibody, where the N-terminal peptide of GPC3 is a secreted form of a peptide found in blood.

Further, the invention relates to the antibody, where the N-terminal peptide of GPC3 is a peptide comprising amino acid residues 1-374 of GPC3 or a peptide comprising amino acid residues 1-358 of GPC3.

Still further, the invention relates to the antibody, which is a monoclonal antibody.

Additionally, the invention relates to the antibody, which is immobilized to an insoluble support.

Still additionally, the invention relates to the antibody, which is labeled with a labeling material.

Still more additionally, the invention relates to an antibody against a C-terminal peptide of GPC3.

Still further, the invention relates to the antibody, where the C-terminal peptide of GPC3 is a peptide comprising amino acid residues 359-580 of GPC3 or a peptide comprising amino acid residues 375-580 of GPC3.

Still further, the invention relates to the antibody, which is a monoclonal antibody.

Additionally, the invention relates to the antibody, which is a chimera antibody.

Additionally, the invention relates to the antibody, which is a cytotoxic antibody.

Still additionally, the invention relates to a cell-disrupting agent comprising the antibody.

Additionally, the invention relates to the cell disrupting agent, where the cell is a cancer cell.

Further, the invention relates to an anti-cancer agent comprising the antibody.

Additionally, the invention relates to a method for inducing cytotoxicity comprising contacting a cell with the antibody.

Still more additionally, the invention relates to the method, where the cell is a cancer cell.

The invention is now described in detail hereinbelow.

The invention provides an antibody against the secreted form of glypican 3 (GPC3), which is capable of detecting the secreted form of GPC3 in a test sample. By detecting the secreted form of GPC3 in vitro in a test sample, it can be diagnosed whether or not the test subject is afflicted with cancer, particularly hepatoma.

Detection includes quantitative or non-quantitative detection, and includes for example a simple assay for the existence of GPC3 protein, an assay for the existence of GPC3 protein at a given amount or more, and a comparative assay for

the amount of GPC3 protein with the amount in other samples (for example, control sample) as a non-quantitative assay; and an assay for the concentration of the GPC3 protein and an assay for the amount of the GPC3 protein as a quantitative assay.

The test sample includes, but is not limited to, any samples possibly containing the GPC3 protein. A sample collected from biological bodies of mammals is preferable. Further, samples collected from humans are more preferable. Specific examples of such test sample include blood, interstitial fluid, plasma, extravascular fluid, cerebrospinal fluid, synovial fluid, pleural fluid, serum, lymphoid fluid, saliva, and urine. Preferably, the test sample is blood, serum or plasma. Additionally, samples obtained from test samples, such as a culture medium of cells collected from biological bodies are also included in the test sample in accordance with the invention.

The cancer to be diagnosed using the antibody against the N-terminal peptide of GPC3 in accordance with the invention includes, but is not limited to, hepatoma, pancreatic cancer, lung cancer, colon cancer, breast cancer, prostate cancer, leukemia, and lymphoma. Preferably, the cancer is hepatoma.

Because the antibody against the C-terminal peptide of GPC3 in accordance with the invention has a high cytotoxic activity, the antibody can be used for disrupting cancer cells, i.e. for therapeutically treating cancer. Cancer possibly

treated clinically using the antibody includes, but is not limited to, hepatoma, pancreatic cancer, lung cancer, colon cancer, breast cancer, prostate cancer, leukemia, and lymphoma. Preferably, the cancer is hepatoma.

1. Preparation of the anti-GPC3 antibody against the N-terminal peptide or the anti-GPC3 antibody against the C-terminal peptide

The amino acid sequence and nucleotide sequence of GPC3 are described in Lage, H. et al., Gene 188 (1997), 151-156 or GenBank: Z37987.

The anti-GPC3 antibody against the N-terminal peptide or the anti-GPC3 antibody against the C-terminal peptide used in the invention should be capable of specifically binding to the N-terminal peptide of the GPC3 protein or the C-terminal peptide of the GPC3 protein, respectively. The origin or type thereof (monoclonal, polyclonal) or the shape thereof is not specifically limited. Specifically, known antibodies such as mouse antibody, rat antibody, human antibody, chimera antibody and humanized antibody can be used.

When GPC3 is cleaved at a cleavage site, the GPC3 is cut into a peptide of about 40 kDa and a peptide of about 30 kDa, which are on the N-terminal side and the C-terminal side, respectively. The cleavage site of GPC3 is the amino acid residue 358, the amino acid residue 374 or a region in the vicinity

thereof. The main cleavage site is believed to be the amino acid residue 358.

The N-terminal peptide of GPC3 is an N-terminal peptide of GPC3 and of about 40 kDa, which is found in the soluble form of the GPC3 core protein. The N-terminal peptide is preferably a peptide of an amino acid sequence comprising from Met 1 to Lys 374, or a peptide of an amino acid sequence comprising from Met 1 to Arg 358. More preferably, the N-terminal peptide is a peptide of an amino acid sequence comprising from Met 1 to Arg 358, because the main cleavage site is predicted to be at the amino acid residue 358. In accordance with the invention, fragments of the N-terminal peptide may also be employed. In this specification, the N-terminal peptide is also referred to as N-terminal fragment or N-terminal peptide fragment.

In other words, the antibody against the N-terminal peptide of GPC3 in accordance with the invention is an antibody recognizing an epitope existing on the N-terminal peptide of the GPC3 protein. The site of the epitope recognized is not specifically limited.

The C-terminal peptide of GPC3 is a C-terminal peptide of GPC3 and of about 30 kDa found in the soluble form of the GPC3 core protein. Based on the cleavage site mentioned above, the C-terminal peptide is preferably a peptide of an amino acid sequence of from Ser 359 to His 580 or a peptide of an amino acid sequence of from Val 375 to His 580. More preferably,

the C-terminal peptide is a peptide of an amino acid sequence comprising from Ser 359 to His 580, because the main cleavage site is presumed to be at the site of the amino acid residue 358. In accordance with the invention, fragments of such C-terminal peptide may also be employed. In this specification, the C-terminal peptide is also referred to C-terminal fragment or C-terminal peptide fragment.

In other words, the antibody against the C-terminal peptide of GPC3 in accordance with the invention is an antibody recognizing an epitope existing on the C-terminal peptide of the GPC3 protein, and the site of the epitope recognized is not limited.

The antibody may be a polyclonal antibody but is preferably a monoclonal antibody.

The anti-GPC3 N-terminal peptide antibody or the anti-GPC3 C-terminal peptide antibody for use in accordance with the invention can be obtained as a polyclonal antibody or a monoclonal antibody, using known techniques. The anti-GPC3 antibody for use in accordance with the invention is preferably a monoclonal antibody derived from mammals. The monoclonal antibody derived from mammals includes those produced by hybridoma, and those generated in hosts transformed with expression vectors carrying the antibody gene by genetic engineering technology.

Hybridoma producing a monoclonal antibody is prepared

essentially using known techniques as follows. An animal is immunized by a conventional immunization method using GPC3 as a sensitizing antigen to obtain an immune cell, which is then fused to a known parent cell by a conventional cell fusion method. Fused cells are screened for monoclonal antibody-generating cells by a conventional screening method.

Specifically, a monoclonal antibody is prepared as follows.

First, GPC3 for use as a sensitizing antigen for obtaining antibody is prepared by expressing the GPC3 (MXR7) gene/amino acid sequence disclosed in Lage, H. et al., Gene 188 (1997), 151-156. Particularly, the gene sequence encoding GPC3 is inserted in a known expression vector to transform an appropriate host cell, then the intended human GPC3 protein is purified from the host cell or a culture supernatant thereof.

Additionally, naturally occurring GPC3 may also be purified and used.

Then, the purified GPC3 protein is used as a sensitizing antigen. The whole GPC3 protein may be used as a sensitizing antigen. Because an antibody against the N-terminal peptide of the GPC3 protein and an antibody against the C-terminal peptide thereof are also induced in this case, the antibody against the N-terminal peptide of the GPC3 protein and the antibody against the C-terminal peptide thereof may be separately selected. Alternatively, a partial N-terminal

peptide of GPC3 or a partial C-terminal peptide thereof may also be used as a sensitizing antigen. In that case, such partial peptide may be obtained by chemical synthesis on the basis of the amino acid sequence of human GPC3 or by inserting a part of the GPC3 gene into an expression vector or by degrading naturally occurring GPC3 with proteases. The part of GPC3 for use as a partial peptide is the N-terminal GPC3 peptide. A smaller peptide fragment containing the epitope in the part may also be used. Further, a C-terminal peptide of GPC3 may be used as a partial peptide, and a smaller peptide fragment containing the epitope in the part may also be used.

Mammals for immunization with a sensitizing antigen are preferably selected, with taking account of the compatibility with parent cells for use in cell fusion. The mammals used for immunization preferably include, but are not limited to, rodents such as mouse, rat, hamster or rabbit or monkey.

For immunization of animals with a sensitizing antigen, known methods may be employed. Generally, for example, a sensitizing antigen is injected intraperitoneally or subcutaneously in mammals. Specifically, a sensitizing antigen is diluted with or suspended in PBS (phosphate-buffered saline) or physiological saline or the like, to an appropriate volume, and mixed with an appropriate volume of conventional adjuvants, such as Freund's complete adjuvant. After emulsification, the emulsified mixture is administered to

mammals several times every 4 to 21 days. Additionally, an appropriate carrier may be used during the immunization with a sensitizing antigen. In case that a partial peptide of a very small molecular weight is to be used as a sensitizing antigen, the partial peptide may preferably be bound to carrier proteins, such as albumin and keyhole limpet hemocyanin upon immunization.

After mammals are immunized as above and the increase in the level of a desired antigen in serum is observed, immune cells are collected from the mammals, which are then subjected to cell fusion. Preferably, the immune cell is splenocyte.

As another parent cell to be fused to the immune cell, mammalian myeloma cell may be used. As the myeloma cell, known various cell lines are preferably used, including for example P3 (P3x63Ag8. 653) (J. Immunol. (1979) 123, 1548-1550), P3x63Ag8U. 1 (Current Topics in Microbiology and Immunology (1978) 81, 1-7), NS-1 (Kohler G. and Milstein, C. Eur. J. Immunol. (1976) 6, 511-519), MPC-11 (Margulies, D. H. et al., Cell (1976) 8, 405-415), SP2/0 (Shulman, M. et al., Nature (1978) 276, 269-270), F0 (de St. Groth, S. F. et al., J. Immunol. Methods (1980) 35, 1-21), S194 (Trowbridge, I. S. J. Exp. Med. (1978) 148, 313-323), and R210 (Galfre, G. et al., Nature (1979) 277, 131-133).

The cell fusion of the immune cell to the myeloma cell is essentially done by known methods, for example the method of Kohler & Milstein et al. (Kohler G. and Milstein C., Methods

Enzymol. (1981) 73, 3-46).

More specifically, the cell fusion is carried out in conventional nutritious culture media in the presence of a cell fusion stimulator. Cell fusion stimulator includes, for example, polyethylene glycol (PEG) and Sendai virus (HVJ). If desired, auxiliary agents such as dimethylsulfoxide can be added and used so as to enhance the fusion efficiency.

The ratio of an immune cell and a myeloma cell to be used can appropriately be determined. For example, an immune cell at a ratio of 1- to 10-fold a myeloma cell is preferable. Culture medium for use in the cell fusion includes, for example, RPMI1640 and MEM, and other conventional culture media suitable for the growth of myeloma cell lines. Further, auxiliary serum agents such as fetal calf serum (FCS) may be used in combination.

The cell fusion can be done by thoroughly mixing predetermined amounts of immune cells and myeloma cells in the culture medium, adding the resulting mixture to a PEG solution (for example, mean molecular weight of about 1,000 to 6,000) preliminarily heated to about 37 °C, generally to a concentration of 30 to 60 w/v %, and subsequently mixing the mixture to allow the intended fusion cell (hybridoma) to be formed.

Subsequently, a cell fusion agent and the like unpreferable for the growth of hybridoma are removed by adding appropriate culture medium sequentially and centrifuging the mixture to discard the supernatant, and repeating the procedures described

above.

The hybridoma thus obtained is selected by culturing in a conventional selective culture medium, such as HAT medium (containing hypoxanthine, aminopterin and thymidine). The culturing in the HAT medium is continued for a sufficient period of time (typically several days to several weeks) for killing cells (non-fused cells) other than the intended hybridoma cell. Then, a conventional limited dilution method is carried out for screening and single cloning of a hybridoma producing the intended antibody.

The screening and the single cloning of the hybridoma may be done by a screening method on the basis of known antigen-antibody reactions. The antigen is bound to carriers such as beads made of polystyrene and the like, or commercially available 96-well microtiter plates, and reacted with a culture supernatant of the hybridoma. After rinsing the carriers, an enzyme-labeled secondary antibody is added to the plate to determine whether an intended antibody reacting with the sensitizing antigen is contained in the culture supernatant. The hybridoma producing the intended antibody can be cloned by limited dilution method. The N-terminal peptide of GPC3 or a fragment thereof or the C-terminal peptide of GPC3 or a fragment thereof may be used as the antigen for screening.

In addition to obtaining hybridoma by immunizing an animal except humans with an antigen, a human antibody may be prepared

by another method. Human lymphocyte is sensitized with GPC3 in vitro and is then fused to myeloma cell with a permanent division potency derived from humans, to obtain a desired human antibody with a binding activity to the N-terminal peptide of GPC3 or the C-terminal peptide of GPC3 (see JP-B-1-59878). Further, a human antibody against the N-terminal peptide of GPC3 or the C-terminal peptide of GPC3 may be obtained by administering GPC3 as an antigen to a transgenic animal bearing all the repertoires of the genes of human antibodies to obtain a cell producing an anti-GPC3 antibody against the N-terminal peptide or a cell producing an anti-GPC3 antibody against the C-terminal peptide, and then immortalizing the cell (see International Publications WO 94/25585, WO 93/12227, WO 92/03918, and WO 94/02602).

The hybridoma producing the monoclonal antibody thus prepared can be subcultured in a conventional culture medium and can be stored in liquid nitrogen for a long period of time.

One method for obtaining the monoclonal antibody from the hybridoma involves culturing the hybridoma by a conventional method and obtaining the monoclonal antibody from a culture supernatant thereof. Another method involves administering the hybridoma to an animal compatible to the hybridoma for proliferation and obtaining the monoclonal antibody in the form of ascites. The former method is suitable for obtaining the antibody at high purity, while the latter method is suitable

for large-scale production of the antibody.

In accordance with the invention, a monoclonal antibody includes a recombinant antibody produced by gene recombinant technology. A recombinant antibody can be generated by cloning the gene of the antibody from the hybridoma, integrating the gene into an appropriate vector, introducing the gene into a host, and allowing the recombinant antibody to be produced by the host (see for example Vandamme, A. M. et al., Eur. J. Biochem. (1990) 192, 767-775, 1990). Specifically, mRNA encoding the variable (V) region of the anti-GPC3 N-terminal peptide or the anti-GPC3 C-terminal peptide is isolated from the hybridoma generating the anti-GPC3 N-terminal peptide antibody or the hybridoma generating the anti-GPC3 C-terminal peptide antibody, respectively. mRNA isolation can be done by known methods. For example, total RNA is prepared by guanidine ultra-centrifugation method (Chirgwin, J. M. et al., Biochemistry (1979) 18, 5294-5299) or AGPC method (Chomczynski, P. et al., Anal. Biochem. (1987) 162, 156-159), from which the intended mRNA is prepared using the mRNA purification kit (manufactured by Pharmacia). Alternatively, mRNA can directly be prepared using QuickPrep mRNA purification kit (manufactured by Pharmacia).

cDNA of the V region of the antibody is synthesized from the resulting mRNA, using reverse transcriptase. cDNA can be synthesized, using AMV Reverse Transcriptase First-strand cDNA

Synthesis Kit (manufactured by Seikagaku Corporation). cDNA can also be synthesized and amplified using 5'-AmpliFinder Race Kit (manufactured by Clontech) and 5'-RACE method using PCR (Frohman, M.A. et al., Proc. Natl. Acad. Sci. USA (1988) 85, 8998-9002; Belyavsky, A. et al., Nucleic Acids Res. (1989) 17, 2919-2932).

The intended DNA fragment is purified from the resulting PCR product and linked to vector DNA. A recombinant vector is prepared from the vector DNA and introduced in *Escherichia coli* and the like to select a colony for preparation of a desired recombinant vector. Subsequently, the nucleotide sequence of the intended DNA can be confirmed by known methods, for example dideoxynucleotide chain termination method.

After DNA encoding the V region of the intended anti-GPC3 N-terminal peptide antibody or the intended anti-GPC3 C-terminal peptide antibody is obtained, the DNA is inserted into an expression vector containing DNA encoding the desired constant region (C region) of the antibody.

So as to produce the anti-GPC3 N-terminal peptide antibody or the anti-GPC3 C-terminal peptide antibody for use in accordance with the invention, the gene of the antibody is introduced into an expression vector such that the gene is expressed under the control of an expression-regulating region, for example enhancer and promoter. Then, a host cell is transformed with the expression vector, to express the antibody.

The gene of the antibody may be expressed by separately inserting DNA encoding the heavy chain (H chain) of the antibody and DNA encoding the light chain (L chain) thereof in expression vectors to simultaneously transform a host cell, or by inserting DNAs encoding the H chain and the L chain in a single expression vector to transform a host cell (see WO 94/11523).

Additionally, not only such host cells but also transgenic animal can be used for generating a recombinant antibody. For example, the gene of the antibody is inserted intermediately into a gene encoding a protein (e.g., goat β casein) generated inherently in milk to prepare a fusion gene. The DNA fragment comprising the fusion gene with the gene of the antibody as inserted therein is injected in a goat embryo, which is introduced in a female goat. The desired antibody is obtained from the milk produced by a transgenic goat born from the goat having received the embryo or a progeny thereof. So as to increase the amount of milk containing the desired antibody as produced by the transgenic goat, hormone may appropriately be administered to the transgenic goat (Ebert, K. M. et al., Bio/Technology (1994) 12, 699-702)

In accordance with the invention, artificially modified recombinant antibodies, for example a chimera antibody (e.g., humanized antibody) may also be used. These modified antibodies can be produced, using existing methods. In case that the antibody of the invention is to be used as an antibody

for therapeutic treatment, the genetic recombinant type antibody is preferably used.

Chimera antibody can be obtained by linking the DNA encoding the V region of the antibody as obtained in the manner described above to DNA encoding the C region of a human antibody, inserting the resulting DNA in an expression vector, and introducing the vector in a host for production of the antibody. Using this existing method, a chimera antibody useful in accordance with the invention can be obtained.

Humanized antibody is also referred to as reshaped human antibody and is prepared by transplanting the complementarity determining region (CDR) of an antibody of mammals except humans, for example mouse, into the complementarity determining region of a human antibody. General genetic recombination techniques thereof are also known in the art (see European Patent Application EP 125023; WO 96/02576).

Specifically, a DNA sequence designed such that the CDR of mouse antibody can be linked to the framework region (FR) of human antibody is synthetically prepared by PCR, using several oligonucleotides prepared in such a manner that the oligonucleotides might have parts overlapped with the terminal regions of both CDR and FR (see the method described in WO 98/13388).

The FR region of human antibody to be linked to CDR is selected such that the CDR can form a good antigen binding site.

If necessary, the amino acids in the FR in the V region of the antibody may be substituted, so that the CDR of the reshaped human antibody may form an appropriate antigen binding site (Sato, K. et al., Cancer Res. (1993) 53, 851-856).

As the C regions of chimera antibody and humanized antibody, those of human antibody are used; for example, C γ 1, C γ 2, C γ 3, and C γ 4 can be used for the H chain, while C κ and C λ can be used for the L chain. So as to improve the stability of the antibody or the production thereof, the C region of human antibody may be modified.

Preferably, the chimera antibody contains a sequence of an antibody derived from mammals except humans in the V region, and contains a sequence derived from a human antibody in the C region.

Humanized antibody comprises the CDR of an antibody derived from mammals except humans, and the FR and C regions derived from a human antibody. Because the antigenicity of chimera antibody such as humanized antibody is reduced in humans, chimera antibody is useful as an active component of a therapeutic agent of the invention.

The antibody for use in accordance with the invention is not only the whole antibody molecule but also a fragment of the antibody or a modified product thereof, including divalent antibody and monovalent antibody, as long as such fragment or such modified product can bind to the GPC3 N-terminal peptide

or the GPC3 C-terminal peptide. For example, the antibody fragment includes Fab, F(ab')₂, Fv, Fab/C having one Fab and complete FC, or single chain Fv (scFv) where Fv of the H chain and the L chain are linked via an appropriate linker.

Specifically, the antibody is treated with enzymes, for example papain and pepsin, to generate antibody fragments. Otherwise, genes encoding these antibody fragments are constructed, introduced in an expression vector and expressed in an appropriate host cell (see for example, Co, M. S. et al., J. Immunol. (1994) 152, 2968-2976; Better, M. & Horwitz, A. H. Methods in Enzymology (1989) 178, 476-496, Academic Press, Inc.; Plueckthun, A. & Skerra, A. Methods in Enzymology (1989) 178, 476-496, Academic Press, Inc.; Lamoyi, E., Methods in Enzymology (1989) 121, 652-663; Rousseaux, J. et al., Methods in Enzymology (1989) 121, 663-669; Bird, R. E. et al., TIBTECH (1991) 9, 132-137).

ScFv can be obtained by linking the V region of the H chain and the V region of the L chain of an antibody. In this scFv, the V region of the H chain and the V region of the L chain are linked together via a linker, preferably a peptide linker (Huston, J. S. et al., Proc. Natl. Acad. Sci. U.S.A. (1988) 85, 5879-5883). The V region of the H chain and the V region of the L chain in scFv may be derived from any antibodies described herein. Any appropriate single-stranded peptide comprising 12 to 19 amino acid residues may be used as the peptide

linker for linking the V regions.

DNA encoding scFv is obtained by first amplifying DNA encoding the H chain or the V region of the H chain and the DNA encoding the L chain or the V region of the L chain by using as a template a portion of DNA encoding all the sequences thereof or a desired amino acid sequence therein and a pair of primers defining both the ends, and then amplifying the DNA with DNA encoding the peptide linker and a pair of primers defined in such a manner that both the ends of the peptide linker may be linked respectively to the H chain and the L chain.

Once the DNA encoding scFv is prepared, an expression vector carrying the DNA and a host transformed with the expression vector can be obtained by conventional methods. scFv can be obtained using the host by conventional methods.

The antibody fragments can be generated by obtaining and expressing the gene in the same manner as described above and allowing a host to produce the fragments. The "antibody" in accordance with the invention includes such antibody fragments.

There may also be used a modified product of the antibody, for example, anti-glypican antibodies conjugated with various molecules such as labeling substances, toxin, and radioactive materials. The "antibody" in accordance with the invention includes these modified antibodies. Such modified antibodies can be obtained by chemical modification of an antibody. Methods for modifying antibodies have already been established

in the art.

Further, the antibody for use in accordance with the invention may be a bispecific antibody. The bispecific antibody may include those having antigen binding sites recognizing different epitopes on the N-terminal peptide of GPC3 or the C-terminal peptide of GPC3. Alternatively, one of the antigen binding sites recognizes the N-terminal peptide of GPC3 or the C-terminal peptide of GPC3, while the other antigen binding site may recognize a labeling substance and the like. Such bispecific antibody can be prepared or obtained by linking HL pairs of two types of antibodies or by fusing hybridomas generating different monoclonal antibodies together to prepare a fusion cell capable of producing a bispecific antibody. Further, such bispecific antibody can be prepared by genetic engineering technique.

In accordance with the invention, an antibody with a modified sugar chain may also be used for the purpose of enhancing cytotoxic activity. Modification technique of the sugar chain of antibody is known in the art (for example, WO 00/61739, WO 02/31140, etc.).

The antibody gene constructed in the manner described above can be expressed and obtained by known methods. In case of a mammalian cell, a conventional useful promoter, the antibody gene to be expressed and poly(A) signal downstream the 3' side thereof are functionally linked for the expression. For

example, the promoter/enhancer includes human cytomegalovirus immediate early promoter/enhancer.

Additionally, the promoter/enhancer for use in the expression of the antibody for use in accordance with the invention includes, for example, virus promoters including retrovirus, polyoma virus, adenovirus and simian virus 40 (SV40)/enhancer or promoters derived from mammalian cells such as human elongation factor Ia (HEFIa)/enhancer.

In case of using SV40 promoter/enhancer, gene expression can readily be done by the method of Mulligan et al. (Nature (1979) 277, 108). In case of using the HEFIa promoter/enhancer, gene expression can readily be done by the method of Mizushima et al. (Nucleic Acids Res. (1990) 18, 5322).

In case of Escherichia coli, a useful conventional promoter, a signal sequence for antibody secretion and an antibody gene to be expressed are functionally linked for expressing the gene. The promoter includes for example lacZ promoter and araB promoter. In case that lacZ promoter is to be used, the gene can be expressed by the method of Ward et al. (Nature (1998), 341, 544-546; FASEB J. (1992) 6, 2422-2427). In case that araB promoter is to be used, the gene can be expressed by the method of Better et al. (Science (1988) 240, 1041-1043).

As the signal sequence for antibody secretion, pelB signal sequence (Lei, S. P. et al. J. Bacteriol. (1987) 169, 4379) may be used when the antibody is generated in the periplasm

of *Escherichia coli*. After the antibody generated in the periplasm is separated, the structure of the antibody is appropriately refolded for use.

As the replication origin, those from SV40, polyoma virus, adenovirus and bovine papilloma virus (BPV) may be used. For amplification of the copy number of the gene in a host cell system, the expression vector may carry a selective marker, for example, aminoglycoside transferase (APH) gene, thymidine kinase (TK) gene, *Escherichia coli* xanthine guanine phosphoribosyl transferase (Ecogpt) gene and dehydrofolate reductase (dhfr) gene.

So as to produce the antibody for use in accordance with the invention, an appropriate expression system, for example eukaryotic cell or prokaryotic cell system can be used. The eukaryotic cell includes for example established animal cell lines such as mammalian cell lines, insect cell lines, fungal cells and yeast cells. The prokaryotic cell includes for example bacterial cells such as *Escherichia coli* cell.

Preferably, the antibody for use in accordance with the invention is expressed in mammalian cells, for example CHO, COS, myeloma, BHK, Vero, and HeLa cell.

The transformed host cell is cultured in vitro or in vivo to produce the intended antibody. The host cell may be cultured by known methods. As the culture medium, for example, DMEM, MEM, RPMI 1640 and IMDM can be used. Auxiliary serum fluid

such as fetal calf serum (FCS) may also be used in combination.

The antibody expressed and generated as described above can be separated from such cells or host animals and can then be purified to homogeneity. The antibody for use in accordance with the invention can be separated and purified using an affinity column. A protein A column includes, for example, Hyper D, POROS, Sepharose F. F. (manufactured by Pharmacia). Additionally, any separation and purification methods generally used for protein may be employed in the invention. For example, chromatography columns other than affinity column, filter, ultrafiltration, salting-out, and dialysis may be used in combination to separate and purify the antibody (Antibodies A Laboratory Manual, Ed. Harlow, David Lane, Cold Spring Harbor Laboratory, 1988).

2. Detection of GPC3

Using the antibody against the N-terminal peptide of GPC3 in accordance with the invention, GPC3 in a test sample can be detected.

GPC3 to be detected using the antibody of the invention includes, but is not limited to, full-length GPC3 and fragments thereof. So as to detect GPC3 fragments, preferably, a fragment of the N-terminal peptide is detected.

The method for detecting the GPC3 protein in a test sample is not specifically limited. The GPC3 protein is preferably

detected by an immunoassay method using the anti-GPC3 N-terminal peptide antibody. The immunoassay method includes, for example, radioimmunoassay, enzyme immunoassay, fluorescent immunoassay, luminescent immunoassay, immunoprecipitation method, immunonephelometry, western blot technique, immunostaining, and immunodiffusion method. Preferably, the immunoassay method is enzyme immunoassay. Particularly preferably, the immunoassay method is enzyme-linked immunosorbent assay (ELISA) (for example, sandwich ELISA). The immunoassay method such as ELISA as described above can be done by a person skilled in the art according to known methods.

General detection methods using the anti-GPC3 N-terminal peptide antibody to detect the GPC3 protein in a test sample involve, for example, immobilizing the anti-GPC3 N-terminal peptide antibody on a support, adding a test sample to the support for incubation to bind the GPC3 protein to the anti-GPC3 N-terminal peptide antibody, rinsing the support and detecting the GPC3 protein bound through the anti-GPC3 N-terminal peptide antibody to the support.

The support for use in accordance with the invention includes, for example, insoluble polysaccharides such as agarose and cellulose, synthetic resins such as silicone resin, polystyrene resin, polyacrylamide resin, nylon resin and polycarbonate resin, and insoluble supports such as glass. These supports can be used in the forms of beads and plates.

In case of beads, a column packed with beads can be used. In case of plates, multi-well plate (for example, 96-well multi-well plate) and biosensor chip can be used. The anti-GPC3 N-terminal peptide antibody can be bound to the support by general methods such as chemical binding and physical adsorption. Such supports are commercially available.

The binding of the anti-GPC3 N-terminal peptide antibody to the GPC3 protein is generally done in buffers. For example, phosphate buffer, Tris buffer, citric acid buffer, borate salt buffer, and carbonate salt buffer may be used as a buffer. Incubation may be carried out under conditions commonly used, for example, 4 °C to ambient temperature for one hour to 24 hours. Rinsing after incubation may be done using any solutions which do not inhibit the binding of the GPC3 protein to the anti-GPC3 N-terminal peptide antibody. For example, buffers containing surfactants such as Tween 20 may be used.

For the method for detecting the GPC3 protein in accordance with the invention, a control sample may be placed in addition to a test sample containing GPC3 protein to be detected. The control sample includes, for example, a negative control sample containing no GPC3 protein or a positive control sample containing the GPC3 protein. In this case, the GPC3 protein in the test sample can be detected by comparison with the results obtained using the negative control sample containing no GPC3 protein and the results obtained using the positive control

sample containing the GPC3 protein. Additionally, a series of control samples having serially varied concentrations are prepared and the results of detection in the individual control samples are obtained in numerical figure to prepare a standard curve. Based on the standard curve, the GPC3 protein contained in the test sample can be determined quantitatively, based on the numerical figure about the test sample.

A preferable embodiment of the detection of the GPC3 protein bound through the anti-GPC3 N-terminal peptide antibody to the support includes a method using the anti-GPC3 N-terminal peptide antibody labeled with a labeling substance.

For example, a test sample is put in contact with the anti-GPC3 antibody immobilized on a support, which is then rinsed, to detect the GPC3 protein using a labeled antibody specifically recognizing the GPC3 protein.

In this case, the anti-GPC3 N-terminal peptide antibody immobilized on the support and anti-GPC3 N-terminal peptide C antibody labeled with a labeling substance may recognize the same epitope of the GPC3 molecule, but preferably recognize different epitopes.

The anti-GPC3 N-terminal peptide antibody can be labeled by generally known methods. Any labeling substances known to a person skilled in the art can be used, including for example fluorescent dye, enzyme, coenzyme, chemiluminescent substance and radioactive substance. Specific examples thereof include

for example radioisotopes (^{32}P , ^{14}C , ^{125}I , ^3H and ^{131}I), fluorescein, rhodamine, dansylchloride, umbelliferone, luciferase, peroxidase, alkaline phosphatase, β -galactosidase, β -glucosidase, horse radish peroxidase, glucoamylase, lysozyme, saccharide oxidase, microperoxidase, and biotin. Preferably, in the case that biotin is used as a labeling substance, avidin bound with enzymes such as alkaline phosphatase is further added after the addition of a biotin-labeled antibody. For binding the anti-GPC3 antibody with a labeling substance, any of the known methods such as glutaraldehyde method, maleimide method, pyridyl disulfide method and periodate method may be used.

Specifically, a solution containing the anti-GPC3 N-terminal peptide antibody is added to a support, such as a plate, to immobilize anti-GPC3 N-terminal peptide antibody. After rinsing the plate, the plate is blocked with for example BSA, so as to prevent non-specific protein binding. After rinsing again, a test sample is added to the plate. After incubation, the plate is rinsed, to which the labeled anti-GPC3 antibody is added. After appropriate incubation, the plate is rinsed and the labeled anti-GPC3 antibody remaining on the plate is detected. The detection can be done by methods known to a person skilled in the art. For example, in case of labeling with a radioactive substance, the detection can be done by a liquid scintillation or a RIA method. In case of labeling with an enzyme, a substrate for the respective enzyme is added to

detect enzymatic substrate changes via for example color development by spectrophotometer. Specific examples of such substrate include

2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)diammonium salt (ABTS), 1,2-phenylenediamine (ortho-phenylenediamine), and 3,3',5,5'-tetramethylbenzidine (TME). In case of labeling with a fluorescent substance, the fluorescent substance can be detected with fluorophotometer.

A particularly preferable embodiment of the method for detecting the GPC3 protein in accordance with the invention involves using anti-GPC3 N-terminal peptide antibody labeled with biotin and avidin.

Specifically, a solution containing anti-GPC3 N-terminal peptide antibody is added to a support such as plate, to immobilize the anti-GPC3 N-terminal peptide antibody. After rinsing the plate, the antibody is blocked with for example BSA to prevent non-specific protein binding. After rinsing again, a test sample is added to the plate. After incubation, the plate is rinsed, and the biotin-labeled anti-GPC3 antibody is added. After appropriate incubation, the plate is rinsed, and avidin conjugated to an enzyme, such as alkaline phosphatase or peroxidase is added. After incubation, the plate is rinsed, a substrate corresponding to each enzyme conjugated to avidin is added, and the GPC3 protein is detected using an enzymatic substrate change as an indicator.

Another embodiment of the method for detecting the GPC3 protein in accordance with the invention involves using a primary antibody specifically recognizing the GPC3 protein and a secondary antibody specifically recognizing the primary antibody.

For example, a test sample is put in contact with the anti-GPC3 N-terminal peptide antibody immobilized on a support. After incubation, the support is rinsed and the GPC3 protein bound to the support after rinsing is detected using a primary anti-GPC3 antibody and a secondary antibody specifically recognizing the primary antibody. In this case, the secondary antibody is preferably labeled with a labeling substance.

Specifically, a solution containing anti-GPC3 N-terminal peptide antibody is added to a support, such as plate, to immobilize the anti-GPC3 N-terminal peptide antibody. After rinsing the plate, the antibody is blocked with for example BSA to prevent non-specific protein binding. After rinsing again, a test sample is added to the plate. After incubation, the plate is rinsed and a primary anti-GPC3 antibody is added. After appropriate incubation, the plate is rinsed and a secondary antibody specifically recognizing the primary antibody is added. After appropriate incubation, the plate is rinsed and the secondary antibody remaining on the plate is detected. The detection of the secondary antibody can be done by the methods described above.

Still another embodiment of the method for detecting the GPC3 protein in accordance with the invention involves using an aggregation reaction. In this method, GPC3 can be detected using a carrier sensitized with the anti-GPC3 N-terminal peptide antibody. Any carriers may be used as the carrier to be sensitized with the antibody, as far as the carrier is insoluble and stable and does not undergo non-specific reaction. For example, latex particle, bentonite, collodion, kaolin and immobilized sheep erythrocyte may be used. Latex particle is preferably used. Latex particles include, for example, polystyrene latex particle, styrene-butadiene copolymer latex particle, and polyvinyltoluene latex particle. Polystyrene latex particle is preferably used. After the sensitized particle is mixed with a sample and agitated for a given period of time, GPC3 can be detected by observing the aggregation under naked eyes since the aggregation level of such particle is higher as the GPC3 antibody is contained at a higher concentration in the sample. Additionally, the turbidity due to the aggregation can be measured with spectrophotometer and the like, to detect GPC3.

Another embodiment of the method for detecting the GPC3 protein in accordance with the invention involves using a biosensor utilizing surface plasmon resonance phenomenon. The biosensor utilizing surface plasmon resonance phenomenon enables the observation of the protein-protein interaction as

surface plasmon resonance signal on real time using a trace amount of protein without labeling. For example, the binding of the GPC3 protein to the anti-GPC3 N-terminal peptide antibody can be detected by using biosensors such as BIAcore (manufactured by Pharmacia). Specifically, a test sample is put in contact with a sensor chip having the anti-GPC3 N-terminal peptide antibody immobilized thereon, and the GPC3 protein bound to the anti-GPC3 N-terminal peptide antibody is detected as the change of the resonance signal.

The detection methods in accordance with the invention may be automated using various automatic laboratory apparatuses, so that a large volume of samples can be tested at a time.

It is an objective of the invention to provide a diagnostic reagent or kit for detecting GPC3 protein in a test sample for cancer diagnosis. The diagnostic reagent or kit contains at least the anti-GPC3 N-terminal peptide antibody. In case that the diagnostic reagent or kit is based on EIA, a carrier for immobilizing the antibody may be contained, or the antibody may be preliminarily bound to a carrier. In case that the diagnostic reagent or kit is based on the aggregation method using carriers such as latex, the reagent or kit may contain a carrier having the antibody adsorbed thereon. Additionally, the kit may appropriately contain, for example, a blocking solution, a reaction solution, a reaction-terminating solution and reagents for treating sample.

3. Disruption of cancer cell using the anti-GPC3 C-terminal peptide antibody and cancer therapy using the same

(1) Determination of antibody activity

The antigen binding activity of the antibody for use in accordance with the invention may be assayed using known techniques (Antibodies A Laboratory Manual. Ed. Harlow, David Lane, Cold Spring Harbor Laboratory, 1988) and an activity of inhibiting the ligand-receptor binding thereof (Harada, A. et al., International Immunology (1993) 5, 681-690).

A method for assaying the antigen binding activity of the anti-GPC3 C-terminal peptide antibody for use in accordance with the invention includes ELISA (enzyme-linked immunosorbent assay), EIA (enzyme immunoassay), RIA (radioimmunoassay) and fluorescent antibody method. In enzyme immunoassay, a sample containing the anti-GPC3 C-terminal peptide antibody, for example a culture supernatant of a cell producing the anti-GPC3 C-terminal peptide antibody or the purified antibody is added to a plate coated with the GPC3 C-terminal peptide. A secondary antibody labeled with an enzyme such as alkali phosphatase is added and the plate is incubated and rinsed, then an enzyme substrate such as p-nitrophenylphosphoric acid is added to measure the absorbance and assess the antigen binding activity.

So as to determine the activity of the antibody for use in accordance with the invention, the neutralization activity

of the anti-GPC3 C-terminal peptide antibody is measured.

(2) Cytotoxicity

For therapeutic purpose, the antibody for use in accordance with the invention preferably has the ADCC activity or the CDC activity as cytotoxicity.

The ADCC activity can be assayed by mixing an effector cell, a target cell and the anti-GPC3 C-terminal peptide antibody together and examining the ADCC level. As the effector cell, cell such as mouse splenocyte and mononuclear cell separated from human peripheral blood or bone marrow can be utilized. As the target cell, a human cell line such as human hepatoma line HuH-7 can be used. The target cells are preliminarily labeled with ^{51}Cr and incubated with the anti-GPC3 C-terminal peptide antibody, then effector cells at an appropriate ratio is added to the target cells and incubated. After incubation, the supernatant is collected to count the radioactivity in the supernatant, to assay the ADCC activity.

Further, the CDC activity can be assayed by mixing the labeled target cell described above with the anti-GPC3 C-terminal peptide antibody, subsequently adding complement, and counting the radioactivity in the supernatant after incubation.

The Fc moiety is needed for the antibody to exert the cytotoxicity. In case that the inhibitor of cell proliferation in accordance with the invention utilizes the cytotoxicity of

the antibody, thus, the anti-GPC3 C-terminal peptide antibody for use in accordance with the invention preferably contains the Fc moiety.

(3) Cell disruption

The anti-GPC3 C-terminal peptide antibody of the invention may also be used for cell disruption, particularly the disruption of cancer cell. Further, the anti-GPC3 C-terminal peptide antibody of the invention can be used as an anticancer agent. Cancers to be therapeutically treated and prevented by the antibody of the invention include, but are not limited to, hepatoma, lung cancer, colon cancer, breast cancer, prostate cancer, pancreatic cancer and lymphoma, preferably Hepatoma.

(4) Administration method and pharmaceutical formulation

The cell disrupting agent or anticancer agent in accordance with the invention is used for the purpose of therapeutically treating or ameliorating diseases caused by abnormal cell growth, particularly cancer.

The effective dose is selected within a range of 0.001 mg to 1,000 mg per 1 kg body weight. Also the effective dose is selected within a range of 0.01 mg to 100,000 mg/body weight per patient. However, the dose of the therapeutic agents containing the anti-GPC3 C-terminal peptide antibody of the invention are not limited to the above doses.

The timing for administering the therapeutic agent of

the invention is either before or after the onset of clinical symptoms of the diseases.

The therapeutic agent comprising the anti-GPC3 C-terminal-peptide antibody in accordance with the invention as an active component can be formulated by a conventional method (Remington's Pharmaceutical Science, latest edition, Mark Publishing Company, Easton, USA), and may also contain pharmaceutically acceptable carriers and additives.

Examples of such carriers and pharmaceutical additives include water, pharmaceutically acceptable organic solvents, collagen, polyvinyl alcohol, polyvinyl pyrrolidone, carboxyvinyl polymer, carboxymethyl cellulose sodium, polyacrylate sodium, sodium alginate, water-soluble dextran, carboxymethyl starch sodium, pectin, methyl cellulose, ethyl cellulose, gum xanthan, gum arabic, casein, agar, polyethylene glycol, diglycerin, glycerin, propylene glycol, vaseline, paraffin, stearyl alcohol, stearic acid, human serum albumin (HSA), mannitol, sorbitol, lactose and surfactants acceptable as pharmaceutical additives.

In practice, an additive or a combination thereof is selected depending on the dosage form of the therapeutic agent of the invention. However, the additive is not limited to those described above. In case that the therapeutic agent is to be used in an injection formulation, the purified anti-GPC3 C-terminal peptide antibody of the invention is dissolved in

a solvent, such as physiological saline, buffers, and glucose solution, and adsorption preventing agents such as Tween 80, Tween 20, gelatin and human serum albumin is added. Alternatively, the therapeutic agent is provided in a freeze-dried form as a dosage form to be dissolved and reconstituted prior to use. As excipients for freeze-drying, for example, sugar alcohols such as mannitol and glucose and sugars may be used.

Brief Description of the Drawings

Fig. 1 shows bar graphs depicting the results of the analysis of GPC3 mRNA expression using Gene Chip, where Fig. 1A depicts GPC3 expression and Fig. 1B depicts the expression of alpha-fetoprotein (AFP). NL, CH, LC, WD, MD and PD on the horizontal axis represent normal liver, inflammatory lesion of hepatitis, lesion of liver cirrhosis, well-differentiated cancer, moderately differentiated cancer and poorly differentiated cancer, respectively.

Fig. 2 shows images of purified soluble GPC3 of heparan sulfate adduct type and the GPC3 core protein, as stained with CBB.

Fig. 3 shows bar graphs depicting the expression of the GPC3 gene in human hepatoma.

Fig. 4 shows the results of western blotting of the soluble form of the core protein using the anti-GPC3 antibody.

Fig. 5 shows the principle of sandwich ELISA using the anti-GPC3 antibody.

Fig. 6 is a graph of the standard curve for the GPC3 sandwich ELISA using M6B1 and M18D4.

Fig. 7 is a schematic view of the GPC3 structure.

Fig. 8 shows combinations of the anti-GPC3 antibodies employed in ELISA.

Fig. 9 is a graph of the standard curve for the GPC3 sandwich ELISA system using various combinations of the anti-GPC3 antibodies.

Fig. 10 shows the assay results of the ADCC activity of the anti-GPC3 antibody.

Fig. 11 shows the assay results of the CDC activity of the anti-GPC3 antibody.

Best Mode for Carrying out the Invention

The invention is now specifically described in the following Examples. However, the invention is not limited by the Examples.

In the Examples described in this specification, the following materials were used.

As expression vectors of the soluble form of GPC3 and the soluble form of the GPC3 core protein, pCXND2 and pCXND3 prepared by integrating the DHFR gene and the neomycin-resistant gene in pCAGGS were used.

DXB11 was purchased from ATCC. For culturing, 5 % FBS (GIBCO BRL CAT# 10099-141, Lot# A0275242/Minimum Essential Medium Alpha medium (α MEM (+)) (GIBCO BRL CAT# 12571-071)/1 % Penicillin-Streptomycin (GIBCO BRL CAT# 15140-122) was used. For selection of stable cell line of DXB11 expressing each protein, 500 μ g/mL Geneticin (GIBCO BRL CAT# 10131-027)/5 % FBS/ α MEM without ribonucleotides and deoxyribonucleotides (GIBCO BRL CAT# 12561-056)(α MEM(-))/PS was used alone or with supplemented with MTX to a final concentration of 25 nM.

HepG2 was purchased from ATCC and maintained in 10 % FBS/Dulbecco's modified Eagle medium (DMEM) (GIBCO BRL CAT# 11995-065)/PS.

The hybridoma was maintained in 10 % FBS/RPMI1640/1 \times HAT media supplement (SIGMA CAT# H-0262)/0.5 \times BM-Condimed H1 Hybridoma cloning supplement (Roche CAT# 1088947).

Example 1

Cloning and expression analysis of human GPC3 (GPC3) cDNA
Cloning of full-length cDNA encoding human glypican 3 (GPC3 hereinafter)

The full-length cDNA encoding human GPC3 was amplified by PCR, using as a template a first strand cDNA prepared from a colon cancer cell line Caco2 by a general method and Advantage 2 kit (Clontech Cat. No. 8430-1). Specifically, 50 μ l of a reaction solution containing Caco2-derived cDNA of 2 μ l, 1 μ l

of a sense primer (SEQ ID NO: 1), 1 µl of an antisense primer (SEQ ID NO: 2), 5 µl of Advantage2 10 × PCR buffer, 8 µl of dNTP mix (1.25 mM) and 1.0 µl of Advantage polymerase Mix was subjected to 35 cycles of 94 °C for one minute, 63 °C for 30 seconds and 68 °C for 3 minutes. The amplified product from the PCR (inserted in TA vector pGEM-T easy using pGEM-T Easy Vector System I (Promega Cat No. A1360)) was sequenced using ABI3100 DNA sequencer to confirm that cDNA encoding the full-length human GPC3 was isolated. The sequence represented by SEQ ID NO: 3 indicates the nucleotide sequence of the human GPC3 gene, while the sequence represented by SEQ ID NO: 4 indicates the amino acid sequence of human GPC3 protein.

SEQ ID NO: 1: GATATC-ATGGCCGGGACCGTGCGCACCGCGT

SEQ ID NO: 2: GCTAGC-TCAGTGCACCAGGAAGAAGAAGCAC

Expression Analysis of human GPC3 mRNA using GeneChip

mRNA expression was analyzed in 24 cases with hepatoma lesions (well-differentiated cancer: WD; moderately differentiated cancer: MD; poorly differentiated cancer: PD), 16 hepatoma cases with non-cancer lesions (hepatitis lesion: CH, cirrhosis lesion : LC), 8 cases with normal liver: NL (informed consent acquired; available from Tokyo University, School of Medicine and Saitama Cancer Center), using GeneChip™ UG-95A Target (Affymetrix). Specifically, total RNA was prepared using ISOGEN (Nippon Gene) from the individual tissues,

from which 15 µg each of total RNA was used for gene expression analysis according to the Expression Analysis Technical Manual (Affymetrix).

As shown in Fig.1, the mRNA expression level of human GPC3 gene (Probe Set ID: 39350_at) was apparently higher in many of the cases compared with the expression in normal liver tissue, despite the differentiation stages of hepatoma. Furthermore, comparison was made with the mRNA expression of alpha-fetoprotein (Probe Set ID: 40114_at) most commonly used as a diagnostic marker of hepatoma currently. It was shown that even in well-differentiated cancer showing almost no such mRNA expression of alpha-fetoprotein, sufficiently enhanced mRNA expression of GPC3 was observed, and that the ratio of the activation of the mRNA expression of GPC3 was higher. Thus, it is considered that GPC3 detection is useful as a diagnostic method of hepatoma at an early stage.

Example 2

Preparation of anti-GPC3 antibody

Preparation of the soluble form of human GPC3

As a material for preparing anti-GPC3 antibody, the soluble form of the GPC3 protein lacking the hydrophobic region on the C-terminal side was prepared.

Using a plasmid DNA containing the complete full-length human GPC3 cDNA supplied from Tokyo University, Advanced

Technology Institute, a plasmid DNA for expressing the soluble form of the GPC3 cDNA was constructed. PCR was conducted using a downstream primer (5'-ATA GAA TTC CAC CAT GGC CGG GAC CGT GCG C-3') (SEQ ID NO: 5) designed to remove the hydrophobic region on the C-terminal side (564-580 amino acid), and an upstream primer (5'-ATA GGA TCC CTT CAG CGG GGA ATG AAC GTT C-3') (SEQ ID NO.6) with the EcoRI recognition sequence and the Kozak's sequence having been added. The resulting PCR fragment (1711 bp) was cloned in pCXND2-Flag. The prepared expression plasmid DNA was introduced in a CHO cell line DXB11. Selection with 500 µg/mL Geneticin resulted in a CHO line highly expressing the soluble form of GPC3.

Using a 1700-cm² roller bottle, the CHO line highly expressing the soluble form of GPC3 was cultured at a large scale, and the culture supernatant was collected for purification. The culture supernatant was applied to DEAE Sepharose Fast Flow (Amersham CAT# 17-0709-01), washed, and eluted with a buffer containing 500 mM NaCl. Subsequently, the product was affinity purified using Anti-Flag M2 agarose affinity gel (SIGMA CAT# A-2220) and eluted with 200 µg/mL Flag peptide. After concentration with Centriprep-10 (Millipore Cat# 4304), the Flag peptide was removed by gel filtration with Superdex 200 HR 10/30 (Amersham CAT# 17-1088-01). Finally, the product was concentrated using DEAE Sepharose Fast Flow column, and eluted with PBS (containing 500 mM NaCl) containing

no Tween 20 for replacement of the buffer.

Preparation of the soluble form of human GPC3 core protein

Using the wild type human GPC3 cDNA as template, cDNA was prepared by assembly PCR, where Ser 495 and Ser 509 were substituted with Ala. A primer was designed in such a fashion that His tag might be added to the C terminus. The resulting cDNA was cloned in pCXND3 vector. The prepared expression plasmid DNA was introduced in a DXB11 line, followed by selection with 500 µg/mL Geneticin, to obtain the CHO line highly expressing the soluble form of the GPC3 core protein.

A large scale cultivation was done with a 1700-cm² roller bottle, and the culture supernatant was collected for purification. The supernatant was applied to Q sepharose Fast Flow (Amersham CAT# 17-0510-01), washed, and eluted with a phosphate buffer containing 500 mM NaCl. Subsequently, the product was affinity purified using Chelating Sepharose Fast Flow (Amersham CAT# 17-0575-01), and eluted with a gradient of 10-150 mM imidazole. Finally, the product was concentrated with Q sepharose Fast Flow and eluted with a phosphate buffer containing 500 mM NaCl.

SDS polyacrylamide gel electrophoresis showed a smear-like band of 50 to 300 kDa and a band of about 40 kDa. Fig.2 shows the results of the electrophoresis. GPC3 is a proteoglycan of 69 kDa and with a heparan sulfate-addition sequence at the C terminus. It was considered that the

smear-like band corresponds to GPC3 modified with heparan sulfate. The results of amino acid sequencing indicated that the band of about 40 kDa had an origin in the N-terminal fragment. Thus, it was anticipated that GPC3 was more or less cleaved.

So as to remove antibodies against heparan sulfate in the following screening for hybridoma, the soluble form of the GPC3 core protein where a heparan sulfate-addition signal sequence Ser 495 and Ser 509 were substituted with Ala. CHO cell line highly expressing the protein was prepared as above, and the culture supernatant was affinity purified utilizing the His-tag. SDS polyacrylamide gel electrophoresis showed three bands of 70 kDa, 40 kDa and 30 kDa. Amino acid sequencing indicated that the band of 30 kDa was the C-terminal fragment of GPC3. The C-terminal fragment starts from serine 359 or from valine 375. Thus, it was anticipated that GPC3 received some enzymatic cleavage. The reason why the band of 30 kDa was not observed in the GPC3 of heparan sulfate-added type was that the fragment formed the smear-like band due to the addition of heparan sulfate. It is a novel finding that GPC3 receives enzymatic cleavage at a specific amino acid sequence, but the biological meaning thereof has not yet been elucidated.

The inventors made an assumption on the basis of the results that GPC3 on the membrane even in hepatoma patients would be cleaved and secreted as the soluble form in blood. Compared with AFP as a hepatoma marker, the expression of the

gene of GPC3 was found higher in hepatoma patients at earlier stages (Fig. 1). So as to examine the possibility as a novel tumor marker with higher clinical utility than that of AFP, an anti-GPC3 antibody was prepared to construct a sandwich ELISA system as described in Example 2 or below.

Preparation of anti-GPC3 antibody

Because the homology of human GPC3 with mouse GPC3 is as high as 94 % at the amino acid levels, it was considered that it might be difficult to obtain the anti-GPC3 antibody by the immunization of normal mouse with human GPC3. Thus, MRL/lpr mouse with autoimmune disease was used as an animal to be immunized. Five MRL/lpr mice (CRL) were immunized with the soluble form of GPC3. For the first immunization, the immunogen protein was adjusted to 100 µg/animal and was then emulsified using FCA (Freund's complete adjuvant (H37 Ra), Difco (3113-60), Becton Dickinson (cat# 231131)), which was then subcutaneously administered to the mice. Two weeks later, the protein was adjusted to 50 µg/animal and emulsified with FIA (Freund's incomplete adjuvant, Difco (0639-60), Becton Dickinson (cat# 263910)) for subcutaneous administration to the mice. At one week interval since then, booster was carried out in total of 5 times. For final booster, the protein was diluted with PBS to 50 µg/animal, which was administered in the caudal vein. By ELISA using an immunoplate coated with the GPC3 core protein, it was confirmed that the serum antibody

titer against GPC3 was saturated. A mouse myeloma cell P3U1 and mouse splenocyte were mixed together to allow for cell fusion in the presence of PEG1500 (Roche Diagnostics, cat# 783641). The resulting mixture was inoculated in a 96-well culture plate. From the next day, hybridoma was selected with the HAT medium, the culture supernatant was screened by ELISA. Positive clones were subjected to monocloning by limited dilution method. The resulted monoclonal was cultured at an enlarged scale and the culture supernatant was collected. The screening by ELISA was done using the binding activity to the GPC3 core protein as a marker to obtain six clones of an anti-GPC3 antibody with a strong binding potency.

The antibody was purified using Hi Trap Protein G HP (Amersham CAT# 17-0404-01). The supernatant from the hybridoma culture was applied directly to a column, washed with a binding buffer (20 mM sodium phosphate, pH 7.0) and eluted with an elution buffer (0.1 M glycine-HCl, pH 2.7). The eluate was collected into a tube containing a neutralization buffer (1 M Tris-HCl, pH 9.0) for immediate neutralization. After antibody fractions were pooled, the resulting pool was dialyzed against 0.05 % Tween 20/PBS overnight and for a whole day for buffer replacement. NaN_3 was added to the purified antibody to 0.02 %. The antibody was stored at 4 °C.

Analysis of anti-GPC3 antibody

The antibody concentration was assayed by mouse IgG sandwich ELISA using goat anti-mouse IgG (gamma) (ZYMED CAT# 62-6600) and alkali phosphatase-goat anti-mouse IgG (gamma) (ZYMED CAT# 62-6622), along with a commercially available purified mouse IgG1 antibody (ZYMED CAT# 02-6100) as a standard.

The isotyping of the anti-GPC3 antibody was done with ImmunoPure Monoclonal Antibody Isotyping Kit II (PIERCE CAT# 37502) by the method according to the attached manual. The results of the isotyping indicated that all of the antibodies were of IgG1 type.

By western blotting using the GPC3 core protein, the epitopes of the anti-GPC3 antibody were classified. The soluble form of the GPC3 core protein was applied to 10 % SDS-PAGE mini (TEFCO CAT# 01-075) at 100 ng/lane for electrophoresis (60 V for 30 min; 120 V for 90 min), and subsequently transferred on Immobilon-P (Millipore CAT# IPVH R85 10) using Trans-Blot SD Semi-Dry Electrophoretic Transfer Cell (BIO-RAD) (15 V for 60 min). After the membrane was gently rinsed with TBS-T (0.05 % Tween 20, TBS), the membrane was shaken with 5 % skim milk-containing TBS-T for one hour (at ambient temperature) or overnight (at 4 °C). After shaking with TBS-T for about 10 minutes, each anti-GPC3 antibody diluted with 1 % skim milk-containing TBS-T to 0.1 to 10 µg/ml was added for one-hour with shaking. The membrane was rinsed with TBS-T (10 minutes × three times) and shaken with HRP-anti-mouse IgG antibody

(Amersham CAT# NA 931) diluted to 1:1000 with 1 % skim milk-containing TBS-T for one hour, and rinsed with TBS-T (10 minutes \times three times). ECL-Plus (Amersham RPN 2132) was used for chromogenic reaction. Hyperfilm ECL (Amersham CAT# RPN 2103K) was used for detection. Fig.4 shows the results of the western blotting analysis. For the classification, it was determined that the antibody reacting with the band of 40 kDa has an epitope at the N terminus, while the antibody reacting with the band of 30 kDa has an epitope at the C terminus. As antibodies recognizing the N-terminal side, M6B1, M18D4, and M19B11 were obtained. As antibodies recognizing the C-terminal side, M3C11, M13B3, and M3B8 were obtained. The results of the analysis using BIACORE indicated that the KD values of the individual antibodies were in the range of from 0.2 to 17.6 nM.

Example 3

Detection of the secreted form of GPC3

Mouse xenograft model

3,000,000 human hepatoma HepG2 cells were transplanted under the abdominal skin in 6-weeks female SCID mice (Fox CHASE C. B-17/Icr-scidJcl, Japan Clair) and nude mice (BALB/cA Jcl-nu, Japan Clair). 53 days later when tumor was sufficiently formed, whole blood was drawn out from the posterior cava of HepG2-transplanted SCID mice #1, 3, and 4. Plasma was prepared

in the presence of EDTA-2Na and aprotinin (Nipro Neotube vacuum blood tube, NIPRO, NT-EA0205) and stored at -20 °C until assay date. In the case of the HepG2-transplanted SCID mouse #2, whole blood was taken 62 days after HepG2 transplantation. In the case of the HepG2-transplanted nude mice #1 and #2, whole blood was taken 66 days after HepG2 transplantation. As a control, plasma was prepared from normal SCID mouse of the same age by the same procedures.

Sandwich ELISA

So as to detect the secreted form of GPC3 in blood, a sandwich ELISA system of GPC3 was constructed. M6B1 was used as an antibody to be coated in a 96-well plate. M18D4 labeled with biotin was used as an antibody detecting GPC3 bound to M6B1. For chromogenic reaction, AMPAK of DAKO was used for achieving high detection sensitivity.

A 96-well immunoplate was coated with the anti-GPC3 antibody diluted with a coating buffer (0.1 M NaHCO₃, pH 9.6, 0.02 w/v % NaN₃) to obtain a concentration of 10 µg/mL, and incubated at 4 °C overnight. On the next day, the plate was rinsed three times with 300 µl/well of rinse buffer (0.05 v/v %, Tween 20, PBS) and 200 µl of dilution buffer (50 mM Tris-HCl, pH 8.1, 1 mM MgCl₂, 150 mM NaCl, 0.05 v/v % Tween 20, 0.02 w/v % NaN₃, 1 w/v % BSA) was added for blocking. After storage for several hours at ambient temperature or at 4 °C overnight, mouse

plasma or the culture supernatant appropriately diluted with a dilution buffer was added and incubated at ambient temperature for one hour. After rinsing with RB at 300 μ l/well three times, the biotin-labeled anti-GPC3 antibody diluted with a dilution buffer to 10 μ g/mL was added, and incubated at ambient temperature for one hour. After rinsing with RB at 300 μ l/well three times, AP-streptoavidin (ZYMED) diluted to 1/1000 with a dilution buffer was added, and incubated at ambient temperature for one hour. After rinsing with the rinse buffer at 300 μ l/well five times, AMPAK (DAKO CAT# K6200) was added for chromogenic reaction according to the attached protocol, and the absorbance was measured with a microplate reader.

For biotinylation of the antibody, Biotin Labeling Kit (CAT# 1 418 165) of Roche was used. A spreadsheet software GlaphPad PRISM (GlaphPad software Inc. ver. 3.0) was used to calculate the concentration of the soluble form of GPC3 in a sample. Fig.5 shows the principle of the sandwich ELISA in this Example.

Using the purified soluble form of GPC3, a standard curve was prepared. Consequently, a system with a detection limit of several nanograms/mL could be constructed. Fig.6 shows a standard curve for the GPC3 sandwich ELISA using M6B1 and M18D4. Using the system, an attempt was made to detect the secreted form of GPC3 in the culture supernatant of HepG2 and the serum of a mouse transplanted with human hepatoma HepG2. The secreted

form of GPC3 was detected in the culture supernatant of HepG2 and the serum of the mouse transplanted with human hepatoma HepG2, while the secreted form of GPC3 was below the detection limit in the control culture medium and the control mouse serum. On a concentration basis of the purified soluble form of GPC3, the soluble form of GPC3 was at 1.2 µg/mL in the culture supernatant of HepG2 and at 23 to 90 ng/mL in the serum of the mouse (Table 1).

Table 1

Assay of the secreted form of GPC3 in the plasma of a mouse transplanted with HepG2 (ng/mL)

	Tumor volume (mm ³)	M6B01(N)-M 1BD4(N)	M19B11(N)- M18D4(N)	M6B1(N)- BioM3C11(C)	M13B3(C)-Bi oM18D4(N)	M13B3(C)-Bi oM3B8(C)
Culture supernatant of HepG2		1190	1736	224	234	<1
HepG2-transplanted SCID mouse #1	2022	65.4	76.9	<10	<10	<10
HepG2-transplanted SCID mouse #2	1706	71.7	94.8	<10	<10	<10
HepG2-transplanted SCID mouse #3	2257	90.3	113.9	<10	<10	<10
HepG2-transplanted SCID mouse #4	2081	87.3	107.3	<10	15.0	<10
HepG2-transplanted nude mouse #1	1994	58.7	53.6	19.7	35.5	102.2
HepG2-transplanted nude mouse #2	190 & 549	22.9	33.6	<10	11.5	40.6
Normal SCID mouse #1	0	<10	<10	<10	<10	<10
Normal SCID mouse #2	0	<10	<10	<10	<10	<10
Normal SCID mouse #3	0	<10	<10	<10	<10	<10

Structure of secreted form of GPC3

It was examined whether or not the blood-secreted GPC3 has the structure of the N-terminal fragment as preliminarily assumed. In case that the secreted form of GPC3 was the N-terminal fragment, it is considered that the secreted form of GPC3 will not be detected by sandwich ELISA with a combination of an antibody recognizing the N terminus and an antibody recognizing the C terminus. Using three types of each antibody recognizing the N-terminal fragment and each antibody recognizing the C-terminal fragment, sandwich ELISA systems with various combinations were constructed. Fig.7 shows the structure of the secreted form of GPC3 and Fig.8 shows combinations of the antibodies. Fig.9 shows a standard curve of the sandwich ELISA. Table 1 shows the assay results. As shown in Table 1, the secreted form of GPC3 was detected at higher values in the culture supernatant of HepG2 and the serum of a mouse transplanted with human hepatoma HepG2 with combinations of antibodies recognizing the N-terminal fragment, while it was detected below the detection limit in many samples from the mice with the systems containing antibodies recognizing the C-terminal fragment. Thus, it was anticipated that the secreted form of GPC3 dominantly comprises the N-terminal fragment. Accordingly, it was suggested that the blood-secreted GPC3 was possibly detected at a high sensitivity by using an antibody against the amino acid sequence comprising

the amino acid residue 1 to the amino acid residue 374 of GPC3.

Example 4

Preparation of anti-GPC3 mouse-human chimera antibody

Using total RNA extracted from a hybridoma producing an antibody capable of binding to human GPC3 (human GPC3-antibody recognizing C-terminus: M3C11, M1E07; human GPC3-antibody recognizing N terminus: M19B11, M18D04, M5B09, M10D02), the cDNA of variable region of the antibody was amplified by RT-PCR. The total RNA was extracted from the hybridoma of 1×10^7 cells, using RNeasy Plant Mini Kits (manufactured by QIAGEN). Using 1 μ g of the total RNA and also using SMART RACE cDNA Amplification Kit (manufactured by CLONTECH), a synthetic oligonucleotide MHC-IgG1 (SEQ ID NO:7) complementary to the mouse IgG1 constant region sequence or a synthetic oligonucleotide kappa (SEQ ID NO:8) complementary to the nucleotide sequence of the mouse κ chain constant region, a 5'-terminal fragment of the gene was amplified. The reverse-transcription was done at 42 °C for one hour and 30 minutes. 50 μ l of the PCR solution contained 5 μ l of 10 \times Advantage 2 PCR Buffer, 5 μ l of 10 \times Universal Primer A Mix, 0.2 mM dNTPs (dATP, dGTP, dCTP, dTTP), 1 μ l of Advantage 2 Polymerase Mix (all manufactured by CLONTECH), 2.5 μ l of the reverse-transcription product, and 10 pmole of the synthetic oligonucleotide MHC-IgG1 or kappa. After the initial temperature at 94 °C for 30 seconds, a cycle of 94 °C for 5 seconds

and 72 °C for 3 minutes was repeated five times; a cycle of 94 °C for 5 seconds, 70 °C for 10 seconds and 72 °C for 3 minutes was repeated five times; and a cycle of 94 °C for 5 seconds, 68 °C for 10 seconds and 72 °C for 3 minutes was repeated 25 times. Finally, the reaction product was heated at 72 °C for 7 minutes. After the individual PCR products were purified from agarose gel using QIAquick Gel Extraction Kit (manufactured by QIAGEN), the products were cloned in pGEM-T Easy vector (manufactured by Promega), and the nucleotide sequence was determined.

Then, the sequences of the variable regions of the H chain and L chain were linked to the constant regions of the human H chain and L chain. PCR was done using a synthetic oligonucleotide complementary to the 5'-terminal nucleotide sequence of the H chain variable region of each antibody and having the Kozak's sequence and a synthetic oligonucleotide complementary to the 3'-terminal nucleotide sequence and having an NheI site. The resulting PCR products were cloned in a pB-CH vector with the human IgG1 constant region inserted in pBluescript KS+ vector (manufactured by TOYOBO). The mouse H chain variable region and the human H chain (γ 1 chain) constant region are linked together via the NheI site. The prepared H chain gene fragment was cloned in an expression vector pCXND3. The scheme of the construction of the vector pCXND3 is described below. So as to divide the gene encoding the antibody H chain

and the vector sequence from DHFR- Δ E-rvH-PM1-f (see WO 92/19759), the vector was digested at the restriction enzyme EcoRI/SmaI sites to recover only the vector sequence. Subsequently, the vector sequence was cloned in EcoRI-NotI-BamHI adaptor (manufactured by Takara Shuzo Co., Ltd.). This vector was designated as pCHO1. A region from pCHO1 expressing the DHFR gene was cloned in pCXN at the restriction enzyme HindIII site (Niwa et al., Gene 1991: 108: 193-200). The resulting vector was designated as pCXND3. The nucleotide sequences of the H chains of the anti-GPC3 mouse-human chimera antibodies (M3C11, M1E07, M19B11, M18D04) contained in each plasmid are shown as SEQ ID NOS: 9, 11, 13 and 15, respectively. The amino acid sequences thereof are shown as SEQ ID NOS: 10, 12, 14, and 16, respectively. Additionally, PCR was done using a synthetic oligonucleotide complementary to the 5'-terminal nucleotide sequence of the L chain variable region of each antibody and having the Kozak's sequence and a synthetic oligonucleotide complementary to the 3'-terminal nucleotide sequence and having a BsiWI site. The resulting PCR products were cloned in a pB-CL vector, where the human kappa chain constant region was preliminarily inserted in pBluescript KS+ vector (manufactured by TOYOBO). The human L chain variable region and the constant region were linked together via the BsiWI site. The prepared L chain gene fragment was cloned in an expression vector pUCAG. The vector pUCAG is a vector prepared by digesting pCXN (Niwa

et al., Gene 1991: 108: 193-200) with restriction enzyme BamHI to obtain a 2.6-kbp fragment, which is then cloned into the restriction enzyme BamHI site of pUC19 vector (manufactured by TOYOBO). The nucleotide sequences of the L chains of the anti-GPC3 mouse-human chimera antibodies (M3C11, M1E07, M19B11, M18D04) contained in each plasmid are shown as SEQ ID NOS: 17, 19, 21 and 23, respectively. The amino acid sequences thereof are shown as SEQ ID NOS: 18, 20, 22 and 24, respectively.

So as to prepare an expression vector of the anti-GPC3 mouse-human chimera antibody, a gene fragment obtained by digesting the pUCAG vector having the L chain gene fragment inserted therein with restriction enzyme HindIII (manufactured by Takara Shuzo Co., Ltd.) was cloned into the restriction enzyme HindIII cleavage site of pCXND3 having the H chain gene inserted therein. The plasmid will express the neomycin-resistant gene, the DHFR gene and the anti-GPC3 mouse-human chimera antibody gene in animal cells.

A CHO-based cell line for stable expression (DG44 line) was prepared as follows. The gene was introduced by electroporation method using Gene PulserII (manufactured by Bio Rad). 25 µg of each expression vector of the anti-GPC3 mouse-human chimera antibody and 0.75 ml of CHO cells (1×10^7 cells/ml) suspended in PBS were mixed together, and cooled on ice for 10 minutes, which was then transferred into a cuvette and received a pulse at 1.5 kV and 25 µFD. After a recovery

time at ambient temperature for 10 minutes, the cells treated by the electroporation were suspended in 40 mL of a CHO-S-SFMII culture medium (manufactured by Invitrogen) containing 1 × HT supplement (manufactured by Invitrogen). A 50-fold dilution was prepared using the same culture medium, and added at 100 µl/well in a 96-well culture plate. After culturing in a CO₂ incubator (5 % CO₂) for 24 hours, Geneticin (manufactured by Invitrogen) was added to 0.5 mg/mL, and continued cultivation for 2 weeks. The IgG in the culture supernatant from the wells of colonies of a Geneticin resistance transformant cell was assayed by the following concentration assay method. A cell line with high productivity was expanded at an enlarged scale. The cell line stably expressing the anti-GPC3 mouse-human chimera antibody was cultured in a large-scale culturing and the culture supernatant was collected.

The IgG concentration in the culture supernatant was assayed by human IgG sandwich ELISA using Goat Anti-human IgG (manufactured by BIOSORCE) and Goat Anti-human IgG alkaline phosphatase conjugated (manufactured by BIOSORCE) and compared with the commercially available purified human IgG (manufactured by Cappel).

Each anti-GPC3 mouse-human chimera antibody was purified using Hi Trap Protein GHP (manufactured by Amersham). A culture supernatant of a CHO cell line producing the anti-GPC3 mouse-human chimera antibody was directly applied to a column

and eluted with elution buffer (0.1 M glycine-HCl, pH 2.7). Eluate was collected into a tube containing a neutralization buffer (1 M Tris-HCl, pH 9.0) for immediate neutralization. Antibody fractions were pooled and dialyzed against 0.05 % Tween 20/PBS overnight and for a whole day to replace the buffer. NaN₃ was added to the purified antibody to 0.02 % and stored at 4 °C.

Example 5

Preparation of a CHO cell line stably expressing the full length GPC3

Human GPC3 cDNA was obtained by digesting pGEM-T Easy vector with the full-length human GPC3 cDNA cloned therein with restriction enzyme EcoRI (manufactured by Takara Shuzo Co., Ltd.) and cloned in an expression vector pCOS2. The scheme of the construction of the vector pCOS2 is described below. So as to divide the gene of the antibody H chain of DHFR-ΔE-rvH-PM1-f (see WO 92/19759) from the vector, the vector was digested at the restriction enzyme EcoRI/SmaI sites, to recover only the vector sequence. Subsequently, the vector sequence was cloned in EcoRI-NotI-BamHI adaptor (manufactured by Takara Shuzo Co., Ltd.). This vector was designated as pCHO1. A region from pCHO1 expressing the DHFR gene was removed, into which the sequence of the neomycin resistant gene in HEF-VH-gyl (Sato et al., Mol. Immunol. 1994: 31: 371-381) was inserted.

The vector was designated as pCOS2.

A cell line stably expressing the full-length human GPC3 was prepared as follows. 10 μ l of the full-length human GPC3 gene-expressing vector and 60 μ l of SuperFect (manufactured by QIAGEN) were mixed together, to form a complex, which was then added to a CHO cell line DXB11 to introduce the gene. After culturing in a CO₂ incubator (5 % CO₂) for 24 hours, α MEM (manufactured by GIBCO BRL) containing Geneticin (manufactured by Invitrogen) to a final concentration of 0.5 mg/mL and 10 % FBS (manufactured by GIBCO BRL) was used to start selection. The resulting Geneticin-resistant colonies were collected and cell cloning was done by limited dilution method. Individual cell clones were solubilized to confirm the expression of the full-length human GPC3 by western blotting using the anti-GPC3 antibody. A cell strain stably expressing human GPC3 was obtained.

Example 6

ADCC assay using PBMC derived from human peripheral blood

(1) Preparation of human PBMC

Peripheral blood was collected from normal subjects with heparinized syringes, and diluted to 2 fold with PBS (-), and overlaid on Ficoll-PaqueTM PLUS (Amersham Pharmacia Biotech AB). This was centrifuged (500 \times g, 30 minutes, 20 $^{\circ}$ C), and collected the intermediate layer as a mononuclear cell fraction. After

rinsing three times, the resulting fraction was suspended in 10 % FBS/RPMI to prepare a human PBMC solution.

(2) Preparation of target cell

HepG2 cell cultured in 10 % FBS/RPMI 1640 culture medium was detached from the dish using trypsin-EDTA (Invitrogen Corp), divided in each well at 1×10^4 cells/well in a U-bottom 96-well plate (Falcon), and cultured for 2 days. After culturing, 5.55 MBq of chromium-51 was added and the cells were incubated in a 5 % CO₂ gas incubator at 37 °C for one hour. The resulting cells were rinsed once with the culture medium, to which 50 µl of 10 % FBS/RPMI 1640 culture medium was added to prepare a target cell.

(3) Chromium release test (ADCC activity)

50 µl of an antibody solution prepared to each concentration was added to the target cell on ice for 15 minutes. Subsequently, 100 µl of a human PBMC solution was added (5×10^5 cells/well), and incubated in a 5 % CO₂ gas incubator at 37 °C for 4 hours. After incubation, the plate was centrifuged and the radioactivity in 100 µl of the culture supernatant was counted with a gamma counter. The specific chromium release ratio was determined by the following formula:

$$\text{Specific chromium release ratio (\%)} = (A-C) \times 100 / (B-C)$$

"A" represents the mean radioactivity value (cpm) in each well; "B" represents the mean radioactivity value (cpm) in a well where 100 µl of aqueous 2 % NP-40 solution (Nonidet P-40,

Code No. 252-23, Nakarai Tesque) and 50 μ l of 10 % FBS/RPMI culture medium were added to the target cell; and "C" represents the mean radioactivity value (cpm) in a well where 150 μ l of 10 % FBS/RPMI culture medium was added to the target cell. The test was done in triplicate to calculate the mean of the ADCC activity (%) and the standard error.

The results are shown in Fig.10. Among the six types of anti-GPC3 chimera antibodies, the antibodies ch.M3C11 and ch.M1E07 recognizing the C terminus exerted the ADCC activity, while the antibodies ch. M19B11, ch. M18D04, ch. M5E09 and ch. M10D02 recognizing the N terminus hardly exerted the ADCC activity. The above results indicate that the ADCC activities of the chimera antibodies depend on the recognition sites of the antibodies. Further, it was expected that the antibodies recognizing the C terminus of GPC3 were possibly useful in clinical applications since the antibodies recognizing the C terminal sides from the cleavage sites exerted the ADCC activity.

Example 7

Assay of compliment-dependent cytotoxic activity (CDC activity)

(1) Preparation of human albumin veronal buffer (HAVB)

12.75 g of NaCl (superior grade; Wako Pure Chemical Industries, Ltd.), 0.5625 g of Na-barbital (superior grade; Wako Pure Chemical Industries, Ltd.), and 0.8625 g of barbital

(superior grade; Wako Pure Chemical Industries, Ltd.) were dissolved in Milli Q water to 200 mL, and autoclaved (121 °C, 20 minutes). 100 mL of autoclaved warm Milli Q water was added. Then, it was confirmed that the resulting mixture was at pH 7.43 (pH 7.5 recommended). This was defined as 5 × Veronal Buffer. 0.2205 g of $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ (superior grade; Wako Pure Chemical Industries, Ltd.) was dissolved in 50 mL of Milli Q water to 0.03 mol/L. The resulting solution was defined as CaCl_2 solution. 1.0165 g of $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$ (superior grade; Wako Pure Chemical Industries, Ltd.) was dissolved in 50 mL of Milli Q water to 0.1 mol/L. The resulting solution was defined as MgCl_2 solution. 100 mL of 5 × Veronal Buffer, 4 mL of human serum albumin (Buminate^R 25 %, 250 mg/mL of human serum albumin concentration, Baxter), 2.5 mL of the CaCl_2 solution, 2.5 mL of the MgCl_2 solution, 0.1 g of KCl (superior grade; Wako Pure Chemical Industries, Ltd.), and 0.5 g of glucose (D(+)-glucose, anhydrous glucose, superior grade; Wako Pure Chemical Industries, Ltd.) were dissolved in Milli Q water to 500 mL. This was defined as HAVB. After filtration and sterilization, the resulting solution was stored at a set temperature of 5 °C.

(2) Preparation of target cell

CHO cell expressing GPC3 on the cell membrane as prepared in Example 4 was cultured in alpha-MEM nucleic acid (+) culture medium (GIBCO) supplemented with 10 % FBS and 0.5 mg/mL Geneticin

(GIBCO), detached from the dish using a cell dissociation buffer (Invitrogen Corp), and divided at 1×10^4 cells/well in each well of a 96-well flat bottom plate (Falcon), for culturing for 3 days. After culturing, 5.55 MBq of chromium-51 was added, and incubated in a 5 % CO₂ gas incubator at 37 °C for one hour. The resulting cell was rinsed twice with HAVB, to which 50 µl of HAVB was added to prepare a target cell.

(3) Chromium release test (CDC activity)

Each chimera antibody was diluted with HAVB to prepare an antibody solution of 40 µg/mL. The antibody solution was added in a 50 µl-portion to the target cell, which was then left on ice for 15 minutes. Subsequently, baby rabbit compliment (Cedarlane) diluted with HAVB was added in 100 µl portions to each well to a final concentration of 30 % (final antibody concentration of 10 µg/mL), and incubated in a 5 % CO₂ gas incubator at 37 °C for 90 minutes. After centrifugation of the plate, a 100-µl portion of the supernatant was recovered from each well, and the radioactivity was measured with a gamma counter. The specific chromium release ratio was determined by the following formula:

$$\text{Specific chromium release ratio (\%)} = (A-C) \times 100 / (B-C)$$

"A" represents the mean radioactivity value (cpm) in each well; "B" represents the mean radioactivity value (cpm) in a well where 100 µl of aqueous 2 % NP-40 solution (Nonidet P-40, Code No. 252-23, Nakarai Tesque) and 50 µl of HAVB were added

to the target cell; and "C" represents the mean radioactivity value (cpm) in a well where 150 μ l of HAVB was added to the target cell. The test was done in triplicate to calculate the mean of the CDC activity (%) and the standard error.

The results are shown in Fig.11. Among the six types of the anti-GPC3 chimera antibodies, the antibodies ch.M3C11 and M1E07 recognizing the C terminus exerted the CDC activity, while the antibodies ch. M19B11, ch. M18D04, ch. M5E09 and ch. M10D02 recognizing the N terminus exerted low CDC activities. The above results indicate that the CDC activities of the chimera antibodies depend on the recognition sites of the antibodies. Further, it was expected that the antibodies recognizing the C terminus of GPC3 were possibly useful in clinical applications since the antibodies recognizing the C terminal sides from the cleavage sites exerted the CDC activity.

Industrial Applicability

As shown in the Examples, it was suggested such that a portion of GPC3 highly expressed in hepatoma cells may exist as a secreted form in blood. Because the gene expression of GPC3 is observed at an earlier stage than that of AFP, a hepatoma marker, GPC3 detection is expected to be useful for cancer diagnosis. It is observed that GPC3 is expressed in cancer cell lines other than hepatoma cell lines, such as lung cancer, colon cancer, breast cancer, prostate cancer, pancreatic cancer

and lymphoma. Accordingly, GPC3 is possibly applicable to the diagnosis of cancers other than hepatoma.

Additionally, it is also suggested that a secreted form of GPC3 in blood predominantly comprises the N-terminal fragment of about 40 kDa, which is observed in the soluble form of the GPC3 core protein. This indicates that antibodies recognizing the N-terminal fragment are useful as the antibody for use in such diagnosis. In addition, if antibodies recognizing the C-terminal fragment with the ADCC activity and/or the CDC activity are used for treating hepatoma, the antibodies can efficiently reach hepatoma cell without being trapped by the secreted form of GPC3 present in blood. Thus, such antibodies are useful as agents for disrupting cancer cells and as anti-cancer agents.

The contents of all the publications listed in this specification are entirely included in the specification. Additionally, a person skilled in the art will readily understand that various modifications and variations of the invention are possible without departure from the technical scope and inventive range described in the attached claims. It is intended that the invention also encompasses such modifications and variations.